

Antioxidant Properties and Cytotoxic Activity of Ethyl Acetate Fraction of *Plectranthus amboinicus* (Lour.) Spreng. Leaves on HeLa and T47D Cell Lines

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Abstract

Research into plants with anticancer effects is actively encouraged in order to discover new drugs with lesser toxicity but more potent effects. The aims of study are to evaluate the antioxidant properties and to investigate the cytotoxic activity of *Plectranthus amboinicus* (Lour.) Spreng. leaves ethyl acetate fractions on HeLa, T47D and MCF7 cell lines. The extract was prepared by graded maceration using n-hexane and ethyl acetate. The ethyl acetate extract was fractionated in vacuum liquid chromatography with n-hexane: ethyl acetate; and ethyl acetate: methanol as mobile phase. Then, the fractions were analyzed with thin layer chromatography (TLC). The free radical scavenging activity was measured by DPPH method, the total flavonoid content was calculated by quercetin equivalent and the absorbance is measured by using UV-Visible spectrophotometry. The cytotoxic activity were determined using MTT assay. The fractions contained 5 sub fractions with same TLC profile. The fractions showed antioxidant activity by DPPH method with different IC₅₀ values, namely: 130 µg/mL (I), 127 µg/mL (II), 137 µg/mL (III), 129 µg/mL (IV), and 124 µg/mL (V), respectively. The measurement of total flavonoid content showed 118 mg QE/g (I), 50 mg QE/g (II), 207 mg QE/g (III), 56 mg QE/g (IV), and 55 mg QE/g (V). The IC₅₀ of each sub fractions on HeLa cell were 77 µg/mL, 46 µg/mL, 93 µg/mL, 71 µg/mL and 476 µg/mL; for T47D cell were 1621 µg/mL, 111 µg/mL, 128 µg/mL, 150 µg/mL and 209 µg/mL; and for MCF7 were 259 µg/mL, 343 µg/mL, 575 µg/mL, 408 µg/mL and 250 µg/mL. Based on the results, the fractions derived from ethyl acetate extract of *Plectranthus amboinicus* (Lour.) Spreng. leaves exhibit antioxidant. The Fraction II from ethyl acetate extract of *Plectranthus amboinicus* (Lour.) Spreng. was the most cytotoxic on HeLa, T47D and MCF7 cell lines. It is potential to undergo further isolation of its cytotoxic compounds.

Keywords : antioxidant, cytotoxic, *Plectranthus amboinicus* (Lour.) Spreng., ethyl acetate fractions

INTRODUCTION

The use of medicinal plant extracts for the treatment of human disease is an ancient practice that has been significantly increasing in recent years. Free radicals trigger the degenerative disease such as cancer. Cancer has become one

of the most prevalent and distressing disease with increasing sufferer in the last 50 years (Asrin,

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et al., 2017; Dalimunthe, *et al.*, 2018). The free radicals can be neutralized by antioxidant from intracellular (endogen antioxidant) and outside the body which comes from plants (Thangavelu, *et al.*, 2015). Research into plants with anticancer effects is actively encouraged in order to discover new drugs with lesser toxicity but more potent effects (Kaewthawee and Brimson, 2013; Elgadir, *et al.*, 2015).

Indonesia has diverse plant species which can be utilized as medicinal plants. One of these medicinal plants is *Plectranthus amboinicus* (Lour.) Spreng. This plant was reported to contain ursolic acid which is a pentacyclic triterpenoid carboxylic acid and pharmacologically active (Shan, *et al.*, 2009; Wang, *et al.*, 2011). It is one of the chemopreventive agents which able to suppress the cancer cell proliferation and induce apoptosis (Gupta, *et al.*, 2015; Kamuhabwa, *et al.*, 2000). Previous studies have showed that the ethyl acetate extract with the highest amount of phenolic compounds exhibited the greatest antioxidant activity (Hasibuan, *et al.*, 2013).

The *in vitro* cytotoxic property of the leaves crude extract was tested against cervical adenocarcinoma (HeLa) cells, and the result displayed cytotoxic effect of n-hexane, ethyl acetate and ethanol extracts on HeLa cells with IC_{50} values 76.322 $\mu\text{g/mL}$, 143.291 $\mu\text{g/mL}$, and 88.997 $\mu\text{g/mL}$, respectively (Rosidah and Hasibuan, 2014). It also showed cytotoxic effect on MCF7 breast cancer cell lines concurrently (Hasibuan, *et al.*, 2013).

The previous studies showed that n-hexane, ethylacetate extracts exhibited strong cytotoxic effect on T47D breast cancer cells with IC_{50} value of 44.716 $\mu\text{g/mL}$ and 37.61 $\mu\text{g/mL}$, respectively and showed the synergistic effect in combination with doxorubicin to inhibit the HeLa cell line (Hasibuan and Rosidah, 2016). It displayed the same effect in combination with doxorubicin to inhibit T47D cell line (Hasibuan, *et al.*, 2015). Thus, the extract has potential effect as a chemoprevention. The aims of this study are to investigate the antioxidant and

cytotoxic activities of the ethyl acetate fractions of *Plectranthus amboinicus* (Lour.) Spreng on HeLa and T47D cell lines.

MATERIALS AND METHODS

Chemicals and reagents

Ethyl acetate, n-hexane were purchased from Merck (Darmstadt, Germany), so does $AlCl_3$ and natrium acetate. Meanwhile 2, 2-Diphenyl-1-picrylhydrazyl (DPPH), 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT), and quercetin were from Sigma Chemical (St. Louis, USA). Silica gel 60H and dimethyl sulfoxide (DMSO) were from Sigma Aldrich Chemie GmbH (Schnelldorf, Germany). RPMI media and Phosphate Buffer Saline (FBS) 10% v/v were from Gibco (New York, USA).

Preparation of extract and fractions

The *Plectranthus amboinicus* was obtained from Pematang Siantar, North Sumatera, Indonesia. The leaves of *Plectranthus amboinicus* were dried at 45°C for 7 days in drying cabinets and ground into powder, then followed by extracted with n-hexane through maceration method for three days at room temperature. The supernatant was separated by decantation and the marc was remacerated twice. The extracted marc was then re-extracted with marc of ethyl acetate by maceration. Extract from each solvent were concentrated by a rotary evaporator (Heidolph VV-200, Sigma Aldrich Chemie GmbH) and the concentrated extract was dried by freeze-dryer (Edwards, London, England). The extract was fractionated with n-hexane, and ethyl acetate with vacuum liquid chromatography by using gradient eluent (100:0; 90:10; 80:20; 70:30; 60:40; 50:50; 40:60; 30:70; 20:80; 10:90; 0:100).

In stationary phase, ethyl acetate fractions was fractionated with gradient eluent ethyl acetate: methanol (100:0; 90:10; 80:20; 70:30; 60:40; 50:50; 40:60; 30:70; 20:80; 10:90; 0:100), methanol (100) and silica gel 60H. All fractions were concentrated by rotary evaporator and were freeze-dried to

eliminate any remaining water. Then, the fractions were analyzed by thin layer chromatography with silica gel GF254 and selected n-hexane-ethyl acetate eluent.

Determination of DPPH scavenging activity

The free radical scavenging activity of *Plectranthus amboinicus* ethyl acetate fraction, and quercetin was measured in terms of hydrogen donating or radical scavenging ability using the stable DPPH (Rosidah, 2008; Atolani and Olantuji, 2016). Each of 7.5 mL; 8.75 mL; 10 mL; 11.25 mL from *Plectranthus amboinicus* extract and each of 0.25 mL; 0.5 mL; 0.75 mL from Quercetin (in methanol) were placed in different test tubes. To this mixture, 5 mL 0.5 mM DPPH was added. After 30 min of incubation at room temperature (22-24°C), absorbance was measured at 517 nm by using spectrophotometer (Shimadzu, Kyoto, Japan) with methanol as the blank. A control contained 1 mL methanol and 5 ml 0.5 mM DPPH. Free radical scavenging activity of the extracts (%) was calculated according to the following formula: $(Ac - As)/Ac \times 100$. Where As is the absorbance of DPPH and sample and Ac is the absorbance of control.

Total flavonoid content (TPC) method

Two mL of ethyl acetate fraction of *Plectranthus amboinicus* (Lour.) Spreng. was blended with 0.1 mL aluminium chloride ($AlCl_3$) 10% reagent (1:10) in the reaction tube. It was mixed and incubated at room temperature for 5 minutes. 0.1 mL natrium asetat (CH_3COONa) and 2.8 mL water was added into the solution, mixed and incubated at room temperature for 60 minutes. The absorption was measured by spectrophotometry UV-Vis at 752 nm. The flavonoid content of ethyl acetate fraction of *Plectranthus amboinicus* (Lour.) Spreng. was calculated by substituting the mean value of sample absorbance to regression equation which was obtained from the calibration curve. The total flavonoid content was presented in milligrams of quercetin per gram of sample (mg QE/g sample)

(Rosidah, *et al.*, 2008; Asrin, *et al.*, 2017; Satria, *et al.*, 2017)

Cell lines and culture conditions

HeLa (cervical adenocarcinoma), T47D (ductal breast adenocarcinoma) and MCF7 (pleural effusion breast adenocarcinoma) cell lines were provided by Paracitology Laboratory, Faculty of Medicine, Universitas Gadjah Mada, Indonesia. The cell lines were cultured in RPMI (Gibco) for HeLa cells and DMEM (Merck) for T47D and MCF7 media, supplemented with 10% (v/v) foetal bovine serum (FBS) (Sigma Aldrich Chemie GmbH), 2% penicillin-streptomycin and 0.5% fungizone (Gibco) in a 37°C incubator with 5% CO_2 .

Cytotoxicity assay

Cytotoxicity was determined by MTT colorimetric assay. Briefly, HeLa, T47D and MCF7 breast cancer cell lines were plated at 10^4 cells/well in a 96-well plate. Each well contained 1×10^4 cells, followed by incubation 24 h at 37°C. Cells were treated with ethyl acetate extract of *Plectranthus amboinicus* (Lour.) Spreng. at various concentrations for 24 hours.

Cells were added with MTT 0.5 mg/mL solution and incubated for 4 hours at 37°C. The reaction was stopped by adding stopper solution containing 10% SDS (Sigma Co, St. Louis, USA) in 0,01 N HCl (Merck, New Jersey, USA) and incubated overnight in room temperature. Absorbance was measured by a plate ELISA reader at λ 595 nm (Bio-rad, California, USA). Percentage of viable cells was calculated from the absorbance data. Percentage of viable cell = $(Absample - Abmedium)/(Abcontrol - Abmedium) \times 100\%$. Where A, B and C are absorbance of control group, treatment group and medium (vehicle), respectively (Hameed, *et al.*, 2016; Illian, *et al.*, 2018).

Statistical analysis

All data was stated in IC_{50} which was analyzed by using probit in regression at SPSS 19. The results were expressed as mean \pm SEM. The test was then

used for statistical analyses with *p*-values of 0.05 and were considered significant.

RESULTS

The result of fractioning test has identified by using thin layer chromatogram were presented in Table 1. These fractions that used as samples for antioxidant and cytotoxic activity test. The antioxidant activity of ethyl acetate fractions of *Plectranthus amboinicus*, (Lour.) Spreng. was identified through the color change of DPPH from purple to yellow; when odd electron from DPPH radicals paired with hydrogen from the extract. The result of 5 concentrations of each ethyl acetate fractions of *Plectranthus amboinicus*, (Lour.) Spreng. with 5th times repeating can be seen on Figure 1.

The result of linear regression and IC₅₀ value of ethyl acetate fractions of *Plectranthus amboinicus*, (Lour.) Spreng. can be seen on Table 2. A substance possesses antioxidant properties when the IC₅₀ value is lower than 200 µg/mL (Molyneux, 2004). From Figure 1 showed that the fractions of ethyl acetate of *Plectranthus amboinicus*, (Lour.) Spreng. have a moderate DPPH scavenging activity.

Quercetin is used as a standard in order to determine the total flavonoid content. The

absorbance value of ethyl acetate fractions of *Plectranthus amboinicus*, (Lour.) Spreng. was plotted against the quercetin curve with correlation coefficient value of 0.99665. The total flavonoid content is expressed in QE (quercetin equivalent), *i.e.*, the amount of milligram quercetin equivalent in 1 gr of sample (Geetha, et al., 2017). The result of total flavonoid content is displayed on Figure 2.

Figure 2 exhibited that fraction III has the highest flavonoid content. Whereas, fraction II has highest IC₅₀ value when compared with the result of DPPH assay. The total flavonoid contents in the fractions described the antioxidant containing in each fractions. TPC was determined by the Folin–Ciocalteu method (Cicco, 2009). The fraction II of *Plectranthus amboinicus* was found to contain low levels of phenolic content 50 mg GAE/g. Phenolic compounds are known as an antioxidant, and they are very important plant constituents because of their free radical scavenging ability due to their hydroxyl groups (Sun and Ho, 2005).

In this study, MTT test was conducted to evaluate the cytotoxic activity of ethyl acetate fractions of *Plectranthus amboinicus*, (Lour.) Spreng. on HeLa, T47D and MCF7 cell lines. The cancer cells were exposed to various concentrations (31.25 - 500 µg/mL) of ethyl acetate fractions for 24 h. As seen on Figure 3, the higher concentration

Table 1. Rf value of *Plectranthus amboinicus*, (Lour.) Spreng. Fraction by using thin layer chromatogram.

Fraction	Moving Phase	Rf value
I	n-hexane : ethyl acetat (100:0)	0.38; 0.4067; 0.5733; 0.667; 0.68
II	n-hexane : ethyl acetat (90:10; 80:20)	0.5733; 0.6667
III	n-hexane : ethyl acetat (70:30; 60:40; 50:50; 40:60; 30:70; 20:80; 10:90)	0.28; 0.8; 0.57; 0.637; 0.68
IV	n-hexane : ethyl acetat (0:100)	0.4; 0.5733; 0.7067
V	ethyl acetat : methanol (100:0; 90:10; 80:20; 70:30; 60:40; 50:50)	0.24; 0.4; 0.5067; 0.5733; 0.6267; 0.68

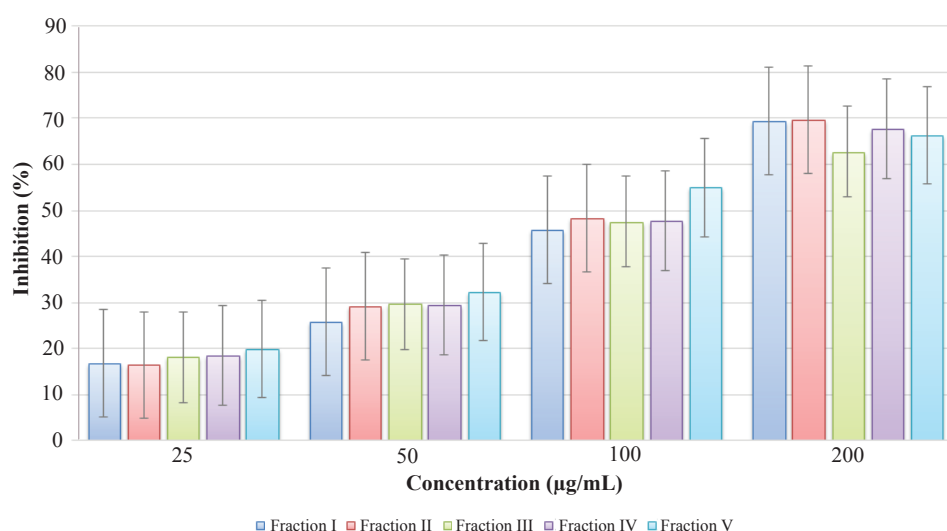


Figure 1. Antioxidant activity of ethyl acetate fractions of *Plectranthus amboinicus*, (Lour.) Spreng. by DPPH method.

of ethyl acetate fractions resulted in decreasing cell viability toward Hela cells. The cytotoxic effect of ethyl acetate fractions of *Plectranthus amboinicus*, (Lour.) Spreng. on T47D cells (Figure 4) showed the similar result to that of Hela cells (Figure 3).

The cytotoxic effect of *Plectranthus amboinicus*, (Lour.) Spreng. fractions on MCF7 cell lines have been shown in Figure 5. According to the results, a non-dependent concentration toxicity was observed in MCF7 cell for all fractions.

This result showed that the value of flavonoid content is not linear with the value of IC_{50} , because not all flavonoid compounds is capable of reducing free radicals which are observed from the differences in the structure of each type.

DISCUSSION

Phenolic compound play an important role in the prevention of cancer disease related to oxidative

Table 2. The antioxidant activity of ethyl acetate fractions of *Plectranthus amboinicus*, (Lour.) Spreng. by DPPH method.

Samples	Regression equation	R value	IC_{50} (µg/mL)
Fraction I	$Y = 0.333x + 6.617$	0.96	130
Fraction II	$Y = 0.335x + 7.632$	0.95	127
Fraction III	$Y = 0.295x + 9.485$	0.91	137
Fraction IV	$Y = 0.320x + 8.700$	0.94	129
Fraction V	$Y = 0.314x + 11.077$	0.88	124
Quercetin	$Y = 9.055x + 5.128$	0.98	4.95

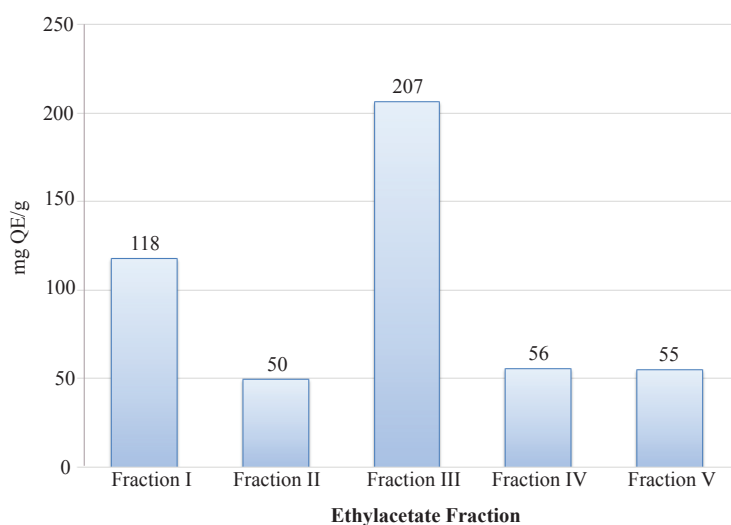


Figure 2. The Flavonoid Content of ethyl acetate fractions of *Plectranthus amboinicus* (Lour.) Spreng. by TPC method.

damage due to their antioxidant properties (Hameed, *et al.*, 2016). The antioxidant activity of phenolic compounds depends on the structure, in particular the number and the positions of the hydroxyl groups and the nature of substitution on the aromatic rings (Middleton, *et al.*, 2000)

Several studies of antioxidant activity show the difference in solvent fractions is closely related

to the total content of phenolic present in them. It is also known that antioxidant activity of a plant extract is not only limited to phenolic and flavonoid but also another compounds, depend on the solvent extraction (Sun and Ho, 2005; Choirunnisa, *et al.*, 2016). Therefore, the relationship between total flavonoid levels and antioxidant activity among plant extracts is complex (Pokorny, *et al.*, 2001).

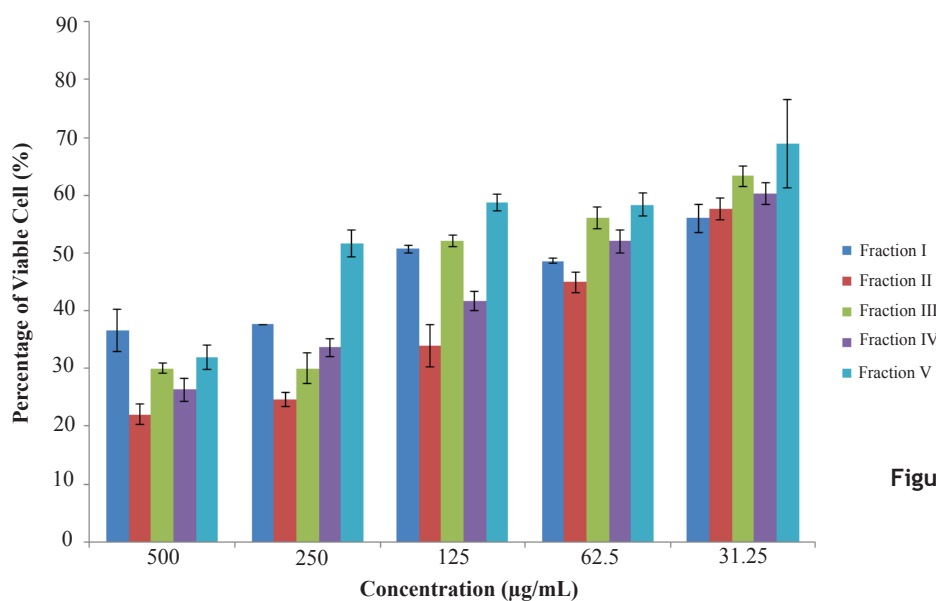


Figure 3. Cytotoxic effect of ethyl acetate fraction of *Plectranthus amboinicus*, (Lour.) Spreng. on HeLa cell line.

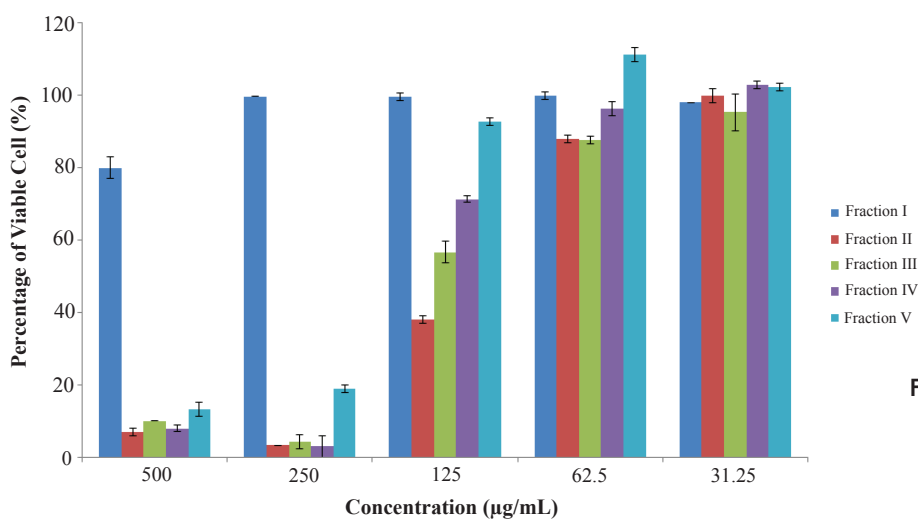


Figure 4. Cytotoxic effect of ethyl acetate fraction of *Plectranthus amboinicus*, (Lour.) Spreng. on T47D cell line.

The cytotoxic activity evaluated by the MTT assays with corresponding IC_{50} are summarized in Table 3. The results showed that the ethyl acetate fractions of *Plectranthus amboinicus*, (Lour.) Spreng. had potent cytotoxic activities. The ethyl acetate fractions performed more potent cytotoxic activities on HeLa than T47D and MCF7 cell lines. Fraction II exhibited significant cytotoxic activity on HeLa, followed by fraction IV, fraction I, and fraction III.

The fractions effect both on T47D and MCF7 cell lines showed unsatisfactory results. The lower IC_{50} values represent the higher potency of the extracts to inhibit the growth of cells. In order to be considered as a potential drug candidate, the IC_{50} value of such agent should be sufficiently low to avoid any possible unspecified effects (Anlar, *et al.*, 2016).

The American National Cancer Institute assigns a significant cytotoxic effect of promising

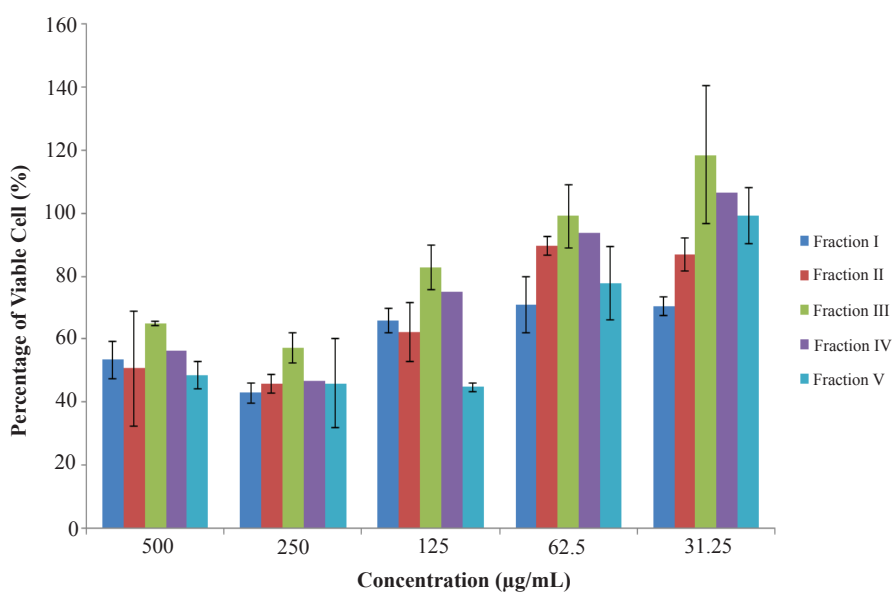


Figure 5. Cytotoxic effect of ethyl acetate fraction of *Plectranthus amboinicus*, (Lour.) Spreng. on MCF7 cell line.

Table 3. Cytotoxic activity (IC₅₀) of ethyl acetate fractions of *Plectranthus amboinicus* (Lour.) Spreng. on HeLa, T47D and MCF7 cell lines.

Fractions	HeLa (µg/mL)	T47D (µg/mL)	MCF7 (µg/mL)
I	77.076	1621.37	259.71
II	46.045	111.19	343.74
III	93.169	127.68	575.11
IV	71.439	149.54	408.54
V	476.009	208.5	250.57

anticancer product for future bio guided studies, which the IC₅₀ value should exert under 30 µg/mL (Hasibuan, et al., 2013). However, according to Kamuhabwa (2000), the potential cytotoxic activity of the extract is less than 100 µg/mL. In this study, the focus is on ethyl acetate fractions, whereas the cytotoxic activity could be due to the presence of active compounds like flavonoid that could probably have inhibitory effects on the cancer cell lines. Several studies have shown high cytotoxic and anticancer activities of flavonoids (Mahadev, et al., 2015). The recent study showed that ethylacetate fraction of *Picria fel-terrae* could inhibit cell grow on G0/G1 phase and induced apoptosis (Satria, et al., 2017). Ethylacetate contained flavonoid which were capable to scavenge the reactive oxygen species effectively because of the phenolic hydroxyl groups and so they are potent antioxidant. Flavonoids reduce breast cancer cell proliferation by inhibiting cell growth, protein kinase activities, and induction of apoptosis (Wang, et al., 2011; Sun, and Ho, 2005).

CONCLUSION

It can be concluded that *Plectranthus amboinicus*, (Lour.) Spreng. ethyl acetate fractions showed DPPH scavenging activity in medium category with the lowest IC₅₀ is 124 µg/mL in fraction V and the highest is 137 ppm in fraction III. The lowest total flavonoid content 50 mg QE/g in fraction II and the highest 207 mg QE/g in

fraction III. Fraction II from ethylacetate extract of *Plectranthus amboinicus*, (Lour.) Spreng. exhibited highest cytotoxic effect both on HeLa and T47D but not on MCF7 cell lines. Fraction V is the most cytotoxic on MCF7 cell line. Required further study to isolate the active compound.

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REFERENCES

- Anlar, H., Bacanli, M., Kutluk, B., Basaran, A.H. and Basaran, N., 2016, Cytotoxic Activity of Resveratrol in Different Cell Lines Evaluated by MTT and NRU Assays, *Turk. J. Pharm. Sci.*, 13(1), 27-34.
- Atolani, O. and Olantuji, G., 2016, Chemical Composition, Antioxidant and Cytotoxic Potential of *Daniellia oliveri* (Rolfe) Hutch. & Dalz, *Turk. J. Pharm. Sci.*, 13(1), 41-46.
- Asrin, H., Hasibuan, P.A.Z. and Marianne, 2017, Total Phenolic Content of Ethanol Extract of *Artocarpus camansi* Leave and Its Effect to Superoxide Dismutase (SOD) Level in Mice, *Indones J. Cancer Chemoprevent.*, 8(3), 101-109.

- Choirunnisa, A.R., Fidrianny, I. and Ruslan, K., 2016, Comparison of Five Antioxidant Assays for Estimating Antioxidant Capacity from Three Solanum Sp Extracts, *Asian J. Pharm. Clin. Res.*, **9**(Supl2), 123-128.
- Cicco, N., Lanorte, M.T., Paraggio, M., Viggiano, M. and Lattanzio, V., 2009, A Reproducible, Rapid, and Inexpensive Folin-ciocalteu Micro-method in Determining Phenolics of Plant Methanol Extracts, *Microchem. J.*, **91**(1), 107-10.
- Dalimunthe, A., Hasibuan, P.A.Z., Silalahi, J., Sinaga, S.F. and Satria, D., 2018, Antioxidant Activity of Alkaloid Compounds from *Litsea cubeba*, Lour, *Orient. J. Chem.*, **34**(2), 1149-1152.
- Elgadir, M. A., Salama, M. and Adam, A.I.S.H.A.H., 2015, Anti-breast Cancer from Various Natural Sources Review, *Int. J. Pharm. Pharm. Sci.*, **7**(2), 44-47.
- Geetha, S., Irulandi, K. and Mehalingam, P., 2017, Evaluation of Antioxidant and Free Radical Scavenging Activities of Different Solvent Extracts of Leaves of *Piper umbellatum*, *Asian J. Pharm. Clin. Res.*, **10**(2), 274-276.
- Gupta, M., Dahiya, J., Marhawa, R.M. and Dureja, H., 2015, Therapies in Cancer Treatment: an Overview, *Int. J. Pharm. Pharm. Sci.*, **7**(4), 1-9.
- Illian, D. N., Basyuni, M., Wati, R. and Hasibuan, P.A.Z., 2018, Polyisoprenoids from *Avicennia marina* and *Avicennia lanata* inhibit WiDr cells proliferation, *Pharmacogn. Mag.*, **14**(58), 513.
- Hasibuan, P.A., Rosidah, Ilyas, S. and Nasution, M.P., 2013, Antioxidant and Cytotoxic Activities of *Plectranthus amboinicus*, (Lour.) Spreng. Extracts, *Int. J. Pharm. Pharm. Res.*, **4**(3), 755-758.
- Hasibuan, P.A.Z. and Rosidah, 2016, Combination Effect of Ethylacetate Extract of *Plectranthus amboinicus*, (Lour.) Spreng. With Doxorubicin Againsts HeLa Cell Lines, *Int. J. Pharm. Clin. Res.*, **8**(5) Suppl, 357-360.
- Hasibuan, PAZ, Chrestella, J. and Satria, D., 2015, Combination Effect of Ethylacetate Extract of *Plectranthus amboinicus*, (Lour.) Spreng. With Doxorubicin Againsts T47D Breast Cancer Cells, *Int. J. Pharm. Pharm. Sci.*, **7**(10), 156-159.
- Hameed, H., Aydin, S., Başaran, A.A. and Başaran, N., 2016, Assessment of Cytotoxic Properties of Sinapic Acid in vitro, *Turk. J. Pharm. Sci.*, **13**(2), 225-232.
- Kamuhabwa, A., Nshimo, C. and de Witte, P., 2000, Cytotoxic of Some Medicinal Plant Extracts Used in Tanzanian Traditional Medicine, *J. Ethnopharmacol.*, **70**(4), 143-149.
- Mahadev, R., Ramakrisnaiah, H., Krishna, V., Deepalakshmi, P. and Kumar, N.N., 2015, Cytotoxic Activity of Methanolic Extracts of *Solanum erianthum*, D. Don, *Int. J. Pharm. Pharm. Sci.*, **7**(2), 106-108.
- Middleton, E., Kandaswami, C. and Theoharides T.C., 2000, The Effect of Plant Flavonoids on Mammalian Cells: Implication for Inflammation, Heart Disease and Cancer, *Pharmacol. Rev.*, **52**(4), 673-751.
- Pokorny, J., Nedyalka, Y. and Michael, G., 2001, *Antioxidants in Food*, England: Woodhead Publishing Limited.
- Rosidah and Hasibuan, P.A.Z., 2014, Cytotoxic Effect of n-Hexane, Ethylacetate, and Ethanol Extracts of *Plectranthus amboinicus*, (Lour.) Spreng. on HeLa and Vero Cells Lines, *Int. J. Pharm. Tech. Res.*, **6**(6), 1806-1809.
- Rosidah, Yam, M.F., Sadikun, A. and Asmawi, M.Z., 2008, Antioxidant Potential of *Gynura procumbens*, *Pharm. Biol.*, **46**(9), 616-625.
- Satria, D., Silalahi, J., Haro, G., Ilyas, S. and Hasibuan, P.A.Z., 2017, Antioxidant and Antiproliferative Activity of Ethylacetate Fraction of *Picria Felterrae*, Lour. Herb, *Asian Pac. J. Cancer Prev.*, **18**(2), 399-403.
- Sitorus, P., Hasibuan, P.A.Z. and Satria, D., 2017, Total Phenolic and Flavonoid Contents and Antioxidant Acitivity of Ethanol Fraction of *Picria felterrae*, Lour. Herbs. *Asian J. Pharm. Clin. Res.*, **10**(7), 243-245.
- Sun, T. and Ho, C., 2005, Antioxidant Activities of Buckwheat Extracts, *Food Chem.*, **90**, 743-749.
- Thangavelu, K., Ravisankar, N., Siddiq, A. and Joseph, J, 2015, In Vitro Antioxidant and Anticancer Potential of Flowers of *Toddalia Asiatica* (Rutaceae), *Int. J. Pharm. Pharm. Sci.*, **7**(3), 95-99.
- Wang, X., Zhang, F., Yang, L. and Mei, Y., 2011, Ursolic Acid Inhibits Proliferation and Induces Apoptosis of Cancer Cells In Vitro and In Vivo, *J. Biomed. Biotechnol.*, **1**, 1-8.